

4 R = R' = R'' = H

5 R' = — Me, R = R'' = H

6 R' = — Me, R = — COMe, R'' = H

7 R' = — Me, R = R'' = — COMe

From these studies it is obvious that compound **1** is the *p*-hydroxybenzoyl ester of the mussaenoside and there is a carboxylic acid group instead of carbomethoxy group at C-4. The *p*-hydroxybenzoyl group is linked at C-2' of the glucose moiety, as shown by the ^{13}C NMR spectra of **1** and its acetate **2**. This was corroborated by the assignments of C-1' (95.9), C-2' (77.3) and C-3' (74.1). The benzoylation effect for C-1', C-2' and C-3', by comparison with the unbenzoylated product, is *ca* -3.5, +3.0, -3.0 respectively. The chemical shift values for the C-8, C-9 and C-10 carbons clearly indicates that the tertiary

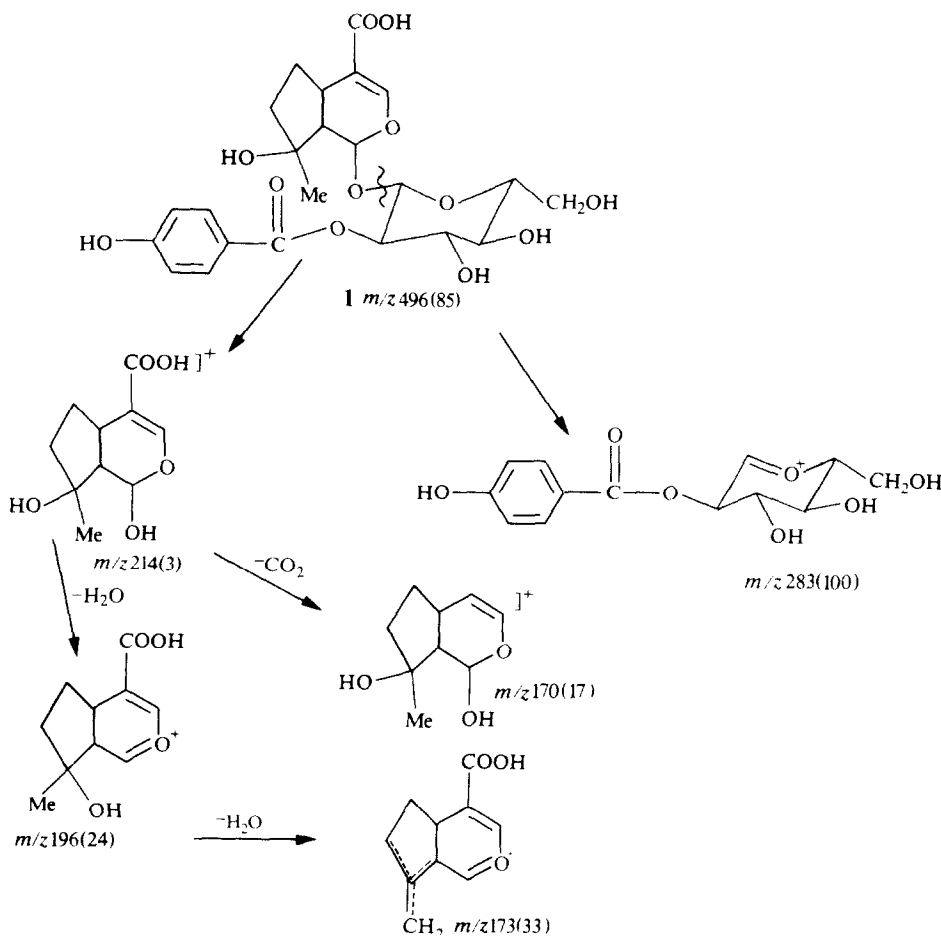
hydroxyl is β at C-8 when compared with other known compounds of similar nature [13].

The structure has been further confirmed by its mass spectral fragmentation pattern which is given in Scheme 1.

EXPERIMENTAL

All mps are uncorr. The leaves of *Vitex negundo* L. (Herbarium no. 11607) were collected locally. The air-dried leaves were first defatted and extracted with CHCl_3 followed by EtOH. The EtOH extract was dried and then subjected to CC over Si gel (2.5 kg) and eluted with EtOAc-MeOH mixtures. Compound **1** was obtained from EtOAc-MeOH (19:1), crystallized from MeOH as white needles, mp 160–162°, $\text{C}_{23}\text{H}_{28}\text{O}_{12}$ $[\alpha]_D^{24}$ -117.6° (MeOH; *c* 3%). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 258; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} : 3300, 1700, 1680, 1640, 1610, 1590, 1430, 1410, 1370, 1310, 1255, 1220, 1065, 1020, 980, 940, 860, 770, 680; ^1H NMR ($\text{DMSO}-d_6$): δ 1.20 (3H, *s*, H-10), 2.20 (1H, *dd*, *J* = 10 Hz, 3.3 Hz, H-9), 5.40 (1H, *d*, *J* = 3.3 Hz, H-1), 6.87 (2H, *d*, *J* = 8.5 Hz, ArH-3'' and H-5''), 7.07 (1H, *d*, *J* = 1.0 Hz, H-3), 7.76 (2H, *d*, *J* = 8.5 Hz, ArH-2'' and 6''). MS *m/z* (rel. int.): 496 [M^+] (85), 478 (10), 283 (100), 196 (24), 180 (5), 178 (33), 170 (17), 152 (3), 138 (4).

Wieffering field test for iridoids [14]. Compound **1** (5 mg) was added to 1 ml of reagent, prepared by mixing HOAc (10 ml) CuSO_4 (1.0 ml of 0.2% soln) and conc HCl (0.5 ml). The mixture was heated over a small flame. After a few seconds a light blue colour was obtained.



Scheme 1.

Table 1. ^{13}C NMR chemical shifts of 2'-*p*-hydroxybenzoyl mussaenosidic acid (1) and its tetra-acetate (2)

C-atom	1	2	Multiplicity
1	93.5	92.8	<i>s</i>
3	148.6	149.5	<i>d</i>
4	112.2	112.6	<i>s</i>
5	29.7	29.4	<i>d</i>
6	28.9	28.4	<i>t</i>
7	41.2	41.5	<i>t</i>
8	77.7	78.9	<i>s</i>
9	50.5	50.9	<i>d</i>
10	24.1	24.0	<i>q</i>
11	167.1*	166.9	<i>s</i>
1'	95.9	95.1	<i>d</i>
2'	77.3	72.3	<i>d</i>
3'	74.1†	70.8	<i>d</i>
4'	70.1	68.3	<i>d</i>
5'	73.1†	72.1	<i>d</i>
6'	60.8	61.6	<i>t</i>
1''	120.5	127.0	<i>s</i>
2''	131.2	131.7	<i>d</i>
3''	114.9	122.3	<i>d</i>
4''	161.5	154.1	<i>d</i>
5''	114.9	122.3	<i>d</i>
6''	131.2	131.7	<i>d</i>
C=O	164.6*	163.8	<i>s</i>
-OCOCH ₃	—	21.1, 20.7, 20.6	<i>q</i>
-OCOCH ₃	—	170.2, 169.4, 170.7, 171.1	<i>s</i>

* and † values are interchangeable.

Acetylation of 1. Treatment of **1** with Ac₂O–pyridine gave the tetra-acetate (**2**) crystallized from EtOAc–petrol as colourless needles mp 120–121°, analysed for C₃₁H₃₆O₁₆. ^1H NMR (CDCl₃): δ 1.30 (3H, *s*, H–10), 2.0–2.20 (9H, 3*s*, 3 × OCOMe), 2.40 (3H, *s*, Ar–OCOMe), 3.8–5.20 (*m*, –CH₂ and CH–OAc of glucose moiety), 5.40 (1H, *d*, *J* = 3.3 Hz, H–1), 7.10 (2H, *d*, *J* = 8.5 Hz, Ar H–3'' and H–5''), 7.30 (1H, *s*, H–3), 8.0 (2H, *d*, *J* = 8.5 Hz, Ar H–2'' and H–6'').

Methylation of 2. Treatment of **2** with CH₂N₂ gave **3** as fine colourless needles from EtOAc–petrol, mp 100–101°, analysed for C₃₂H₃₈O₁₆. ^1H NMR (CDCl₃): δ 1.3 (3H, *s*, H–10), 1.90–2.20 (9H, 3*s*, 3 × OCOMe), 3.33 (3H, *s*, –COOMe), 5.40 (1H, *d*, *J* = 3.3 Hz, H–1), 7.10 (2H, *d*, *J* = 8.5 Hz, Ar H–3'' and H–5''), 7.3 (1H, *s*, H–3), 7.9 (2H, *d*, *J* = 8.5 Hz, Ar H–2'' and H–6'').

Alkaline hydrolysis of 1. Compound **1** (80 mg) was refluxed with methanolic KOH for 30 min. After usual work-up it gave a mixture of two compounds. Compound **4** (30 mg) was separated by CC over Si gel as a colourless powder, analysed for C₁₆H₂₄O₁₀.

Methylation of 4. Treatment of **4** with CH₂N₂ gave a white powder (**5**), analysed for C₁₇H₂₆O₁₀: $[\alpha]_D^{25}$ –114° (MeOH; *c* 0.9%). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 238; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 3400, 1695, 1640; ^1H NMR (DMSO-*d*₆): δ 1.20 (3H, *s*, *ter*–Me), 2.20 (1H, *dd*, *J* = 10.0 Hz and 3.3 Hz, H–9); 3.63 (3H, *s*, COOMe), 5.40 (1H, *d*, *J* = 3.3 Hz, H–1), 7.30 (1H, *d*, *J* = 1.0 Hz, H–3).

Acetylation of 5. Compound **5** with Ac₂O–pyridine gave a

tetra-acetate (**6**) crystallized from MeOH mp 124–126°, analysed for C₂₅H₃₄O₁₄. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 237; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 1750, 1705, 1640; ^1H NMR (CDCl₃): δ 1.33 (3H, *s*, H–10), 1.92–2.09 (12H, 4 × OCOMe), 2.31 (1H, *dd*, *J* = 9.5 Hz, and 3.0 Hz, H–9), 3.03 (1H, *m*, H–5), 3.70 (3H, *s*, COOMe), 5.33 (1H, *d*, *J* = 3.0 Hz, H–1), 7.34 (1H, *d*, *J* = 1.0 Hz, H–3).

Treatment of 6 with Ac₂O–BF₃. To a soln of **6** (40 mg) in Ac₂O was added BF₃–etherate (5 drops) and the mixture was allowed to stand at room temp. for 2 min. Ice-cold H₂O was added and the reaction mixture was extracted with CHCl₃. The organic layer was washed with H₂O, dried over Na₂SO₄, conc and **7** (40 mg) was crystallized from EtOH, as colourless needles mp 116–118°. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 240; IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{–1}: 1745, 1710, 1660; ^1H NMR (CDCl₃): δ 1.51 (3H, *s*, H–10), 1.90–2.08 (3H, *s*, 5 × OCOMe), 2.66 (1H, *dd*, *J* = 8.5 Hz and 2.0 Hz, H–9), 2.95 (1H, *m*, H–5), 3.71 (3H, *s*, COOMe), 5.71 (1H, *d*, *J* = 2.0 Hz, H–1), 7.40 (1H, *d*, *J* = 1.0 Hz, H–3). Analysed for C₂₇H₃₆O₁₅.

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REFERENCES

1. (1976) *The Wealth of India (Raw Materials)* Vol. 10, p. 552. Publication and Information Directorate CSIR, New Delhi.
2. Basu, N. K. and Singh, G. B. (1947) *Q. J. Pharm. Pharmacol.* **20**, 136; *Chem. Abstr.* (1948) 1025.
3. Ghose, T. P. and Krishna, S. (1936) *J. Indian Chem. Soc.* **13**, 634.
4. Banerji, A., Chadha, M. S. and Malshet, V. G. (1969) *Phytochemistry* **8**, 511.
5. Basu, M. M., Ray, G. K. and De, M. K. (1947) *J. Indian Chem. Soc.* **24**, 358.
6. Joshi, V., Merchant, J. R., Nadakarni, Y. V., Namboori, N. and Vaghani, D. D. (1947) *Indian J. Chem.* **12**, 226.
7. Gupta, G. S. and Sharma, D. P. (1973) *Proc. Natl. Acad. Sci. India* **43**, 268; *Chem. Abstr.* (1975) **83**, 128750a.
8. Rao Kodander, U., Rao Venkata, E. and Rao Venkata D. (1977) *Indian J. Pharm.* **39**, 41.
9. Prema, M. and Mishra, G. S. (1978) *Indian J. Chem.* **16B**, 615.
10. Gupta, G. S. and Behari Mukat (1976) *Agra Univ. J. Res. Sci.* **25**, 63; *Chem. Abstr.* (1979) **90**, 51414d.
11. Mishra, G. S. and Subramanian, P. M. (1980) *Planta Med.* **38**, 155.
12. Takeda, Y., Nishimura, H. and Inonye, H. (1977) *Phytochemistry* **16**, 1401.
13. Chaudhuri, R. K., Affi-Yazar, F. U., Sticher, O. and Winkler, T. (1980) *Tetrahedron* **36**, 2317.
14. Trim, A. R. and Hill, R. (1952) *J. Biochem.* **50**, 310.